

# CHITOSAN APPLICATION IN FOOD TECHNOLOGY: A REVIEW OF RECENT ADVANCES

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## ABSTRACT

The review focused on global trends in the development of scientific research and the practical applications of chitosan in food technology in recent years. Chitin and its derivative chitosan obtained from the crustacean shells and the cell wall of fungi are among the most common biopolymers in the world. Chitosan is a polysaccharide discerned by a large number of unsubstituted amino groups. Featured properties of chitosan providing its high chemical and biological activities. Chitosan has various abilities as polycationite, film former, antimicrobial and antioxidant agent. Multifunctional properties open up broad prospects for the chitosan applications in various fields of technology, medicine and industry. The most attention in the review is paid to the works on extending food products shelf life with chitosan based primary edible film coatings and biodegradable packaging. At the same time chitosan applications as an emulsifier, a flocculant, as well as functional food additive, nutrient encapsulating material and dietary supplement are highlighted.

## 1. Introduction

Chitosan — poly- $\beta$ -(1,4)-d-glucosamine is a deacetylated product of chitin — poly- $\beta$ -(1,4)-N-acetyl-d-glucosamine, one of the most abundant natural polysaccharide, found in composition of crustacean shells and fungal cell walls [1]. There is great scientific and practical interest in chitin and chitosan caused to the unique properties as biocompatibility, safety, biodegradability, sorption performance of heavy metals and radionuclides [2]. Scientific publications amount related to chitin and chitosan is increasing annually and several scientific papers related to its production and application are published weekly according to the Web of Science and Google Scholar. Chitin global production in 2015 was 28 thousand tons, and its still increasing [3]. Due to its properties, chitosan has been widely used in various fields of technology, medicine and food industry. Chitosan properties are significantly influenced by its molecular weight (MW), which occurs in a range from 2 to more than 200 kDa. Deacetylation degree (DD) is other important chitosan property affecting an acid solubility which differs it from chitin. Chitosan with DD more than 55% becomes soluble in a 1% acetic or hydrochloric acids [4]. Solubility associated with the protonation of the amino group at the C-2 position of the d-glucosamine unit converts the polysaccharide to a polyelectrolyte under acidic conditions [5].

The purpose of present work is the development progress problem study on food technology chitosan applications and analysis of these trends.

## 2. Main part

A biological activity of chitosan along with safety for the human brings its wide application for food industry. Antioxidant and antimicrobial activity of chitosan allows to use it for increasing the foods shelf life, excellent emulsifying properties gives an opportunity to replace synthetic surfactants in food technologies, also chitosan can be used as a functional ingredient against hypercholesterolemia, hypertension and inflammations, and for the encapsulation of nutrients for a functional food development [6]. Polyelectrolyte and flocculant behavior of chitosan allow to use it as an effective clarifying agent for beverages, as well as for preliminary purification of drinking water [7].

### *Chitosan applications for food shelf life prolongation*

Most significant factors limiting the shelf life of food products are an oxidative and microbial spoilage. Antioxidant activity of

chitosan due to scavenging effect on free radicals. This ability depends on DD and increases with unsubstituted amino groups amount increased. High MW chitosan exhibits a less pronounced antioxidant effect, because its structure is more compact due to the formation of strong intramolecular hydrogen bonds [6]. The antioxidant activities of chitosan in lard and rapeseed oil is comparable to ascorbic acid [8]. Also an antioxidant effect of chitosan in semi-finished products from meat, fish and seafood is noted [9].

The antimicrobial and antifungal activity of chitosan caused by cationite properties appeared in an acidic media. At pH values below  $pK_a$  of chitosan protonated amino groups binding negatively charged carboxyl groups of lipopolysaccharides and peptidoglycans at the bacterial cell wall surface, leading to permeabilization and destruction of external membranes [10]. The mechanism of antifungal action of chitosan is similar to antibacterial and appears to fungi containing a significant amount of polyunsaturated fatty acids [11]. Processing with chitosan solutions of fresh or chopped fruits, berries and vegetables extends their shelf life, inhibits the development of microorganisms and molds. Chitosan addition inhibits bacterial growth in noodle dough, hummus, and cheese [9,12].

Edible film coatings are the most promising chitosan application for food shelf life extending. Edible films are defined as the primary packaging designed to protect foods against external impacts, also it can be used as carrier of active ingredients such as preservatives, flavors and colorants. [13]. Chitosan film coatings are prepared as a solution in dilute acids. Depending on DD, solution concentration, additional components and the drying conditions, there is a wide variety to change physicochemical and biological properties of the resulting compound [14]. Edible coatings reducing weight loss during storage, prevent oxygen permeability, shows antimicrobial and antioxidant activities to, increase the shelf life of processed foods significantly.

The tensile strength of chitosan films produced with acetic acid is higher in comparison with citric, lactic, and malic acids and increases during long-term storage [15]. Elasticity of the coatings could be increased by addition of glycerol or sorbitol as plasticizer. Chitosan-based film coatings effectively extend the shelf life of carrots [16, 17], tomatoes [18], cucumbers [19], bell peppers [20], bananas [21], apples [22], pomegranates [23], pears [24], plums [25], strawberries [26], papaya [27], mango [28], fish [29], meat [30] and minced meat [31, 32].

A number of studies concerning coatings composed of chitosan blends with other film-forming biopolymers allowed to keep biodegradability, non-toxicity, compatibility with food products and improve functional properties. Chitosan-starch blend films shows good water vapor permeability and a high antioxidant activity [33,34]. The addition of cellulose and its derivatives to chitosan films increases elasticity, tensile strength, and optical transparency while maintaining high antimicrobial activity [35,36]. Chitosan-alginate multilayer coatings applied to fresh cut fruits allows to save succulence during a longer while [37,38]. Pectin and chitosan combination occurring a strong intermolecular electrostatic interaction, and the films obtained are characterized by high transparency, strength, and elasticity [39]. Compositions of chitosan with proteins, such as casein and gelatin [40,41,42], are characterized by good mechanical properties, low vapor permeability, and UV-protection due to absorption by peptide bonds [43]. Soy protein or beeswax included to coating composition increasing film hydrophobicity [44,45].

Because of active functional groups chitosan can be used as part of composite materials containing not natural only but synthetic polymer compounds [46]. *Opuntia mucilage* — a complex mixture of functional polysaccharide residues used as film-forming agent with polyvinyl alcohol and chitosan obtaining edible coatings with different properties when the composition is varied. These coatings are hydrophilic due to the *opuntia mucilage*. Inclusion of chitosan has technological approach because it is able to stabilize the film-forming solution increasing zeta potential and also a functional since films with higher content of chitosan became tougher. When increasing polyvinyl alcohol amount film flexibility is increased. That make those mixtures compatible with different food applications required [47]. Films formed by chitosan and polyvinyl alcohol with lignin nanoparticles are characterized by increased strength compared to films formed by individual components, antibacterial action against gram-negative microorganisms and the synergistic antioxidant effect of chitosan and lignin [48].

Natural plant extracts included with the chitosan coating composition has a significant positive effect on the mechanical and biological properties of the protective edible films formed. For example, polyphenols of thyme extract increase tensile strength of chitosan-based films as crosslinkers, and also increase the antioxidant activity [49], turmeric extract addition reduces the UV-permeability, significantly increases the tensile strength and antimicrobial activity [50], milk thistle extract reduces films solubility and water vapor permeability, as well as improves antioxidant properties [51]. Barberry seed extract increases the hydrophobicity of coatings, while whole fruit extract reduces it compared to a control sample. The antimicrobial activity of chitosan films with barberry extracts based quorum sense inhibitory property [52]. Ethanolic extracts of rosemary, cinnamon, guarana and peumus added to chitosan and gelatin based films allows to achieve very high antioxidant and antibacterial activities [53], grapefruit seed extract increases the antifungal activity, elasticity and elongation at break of chitosan films [54].

A significant synergistic antimicrobial effect is observed for composite coatings based on the chitosan matrix including nanoparticles of inorganic antimicrobial agents such as silver [55,56,57] and zinc oxide [58,59,60].

Essential oils addition to film-forming composition gives for the edible films supplemental functional properties. Essential oils increasing antioxidant ability and activity against molds and microorganisms and also changing physical properties of the chitosan films, such as strength, moisture resistance and transparency. That coatings can be used for berries, fruits, fish or meat. Combined action of essential oils and chitosan may lead to synergistic effects. Rosemary oil containing chitosan coating on rainbow trout fillets exhibits more pronounced antibacterial and

antioxidant effect than these components separately [61]. Chitosan coating containing eucalyptus essential oil and  $\alpha$ -tocopherol prolongs the shelf life and reduces fat oxidation of fresh silver carp, which is more related to the combined presence of chitosan and eucalyptus oil than  $\alpha$ -tocopherol used as an regular antioxidant food additive [62]. Cinnamon and clove essential oils enriched chitosan-arabic gum films enhances moisture resistance as well as antimicrobial activity against *Escherichia coli* and *Staphylococcus aureus* more in comparison with the inclusion of these oils by itself [63]. In chitosan-carboxymethylcellulose edible coating cinnamon oil exhibits better antifungal activity add reduces the water vapor permeability by increasing the hydrophobicity than ginger oil but in conjunction emerging a significant synergistic antifungal and moisture resistance effect [64]. Addition of cinnamon and ginger essential oils slightly reduces the transparency of chitosan films, but allows to obtain a thinner coatings. That films effectively prevent caused by fat oxidation and the action of microorganisms fresh pork spoilage [65]. In some cases, essential oils addition to the coating may not have any effect or even negatively affecting on the resulting products. The synergistic antifungal effect on *Penicillium digitatum*, the main cause of citrus green mold when cinnamon essential oil incorporated chitosan coating, showed in vitro was not confirmed when applied to the mandarins [66]. Oregano and basil essential oils reduce tensile strength, while increasing transparency and improving the antimicrobial and antioxidant properties of chitosan-based films [67]. Thyme oil with chitosan coating reduces moisture loss, retains color and inhibits yeast growth, but does not affect on enterobacteria, mesophilic and lactic acid bacteria for ham [68]. Cumin and eucalyptus essential oils as part of the chitosan coating of chicken breasts reducing spoilage caused by various types of microorganisms, extending shelf life and enhancing, the taste and aroma of fried chicken [69]. Lemon oil compared to oregano oil in the chitosan coating composition shows a more pronounced bactericidal effect, both have not adversely affect chilled chicken breast organoleptic [70]. However, lemon oil added with a strawberry coating despite an antifungal resistance increasing negatively changing a typical aroma due to terpenes contained in lemon oil [71].

A promising direction of food quality monitoring is visual edible film colorimetric indicators that change color when the pH accompanying spoilage changes. That coatings were developed for fresh fish on the chitosan-starch base containing red cabbage extract [72], and as polyvinyl alcohol films including chitosan nanoparticles and mulberry extract [73].

Development of biodegradable polymer packaging materials is one of modern trends to extend food shelf life. A microcrystalline cellulose-polyvinyl alcohol-chitosan composition showed two orders of magnitude higher water vapor permeability with comparable strength to polyethylene films and relative elongation exceeds cellophane because of plasticizing effect of polyvinyl alcohol [74]. Polylactic acid with chitosan nanoparticles using polyvinyl alcohol as a plasticizer and polyethylene glycol as crosslinking agent is characterized by a significant increase in tensile strength and antimicrobial activity against aerobic microorganisms [75]. Amphiphilic nanoparticles formed by a chitosan grafted polylactic acid oligomers used as a filler for polylactic acid makes resulting material more flexible and significantly increase the oxygen barrier properties [76].

#### *Chitosan emulsifying applications*

Chitosan is an excellent emulsifier to stabilize heterogeneous oil-in-water systems. Chitosan increases viscosity of the continuous phase, complicating the diffusion of dispersed particles and reducing the droplet aggregation rate. In addition, positively charged amino groups, making chitosan an amphiphilic surfactant polymer.

Chitosan can be used as a single emulsifier, however the emulsions obtained are reversible due to the sensitivity to the pH values. In an acidic media with  $\text{pH} < \text{pK}_a$  protonated chitosan exhibits surface-active properties, interacting with triglycerides carbonyl groups. With pH increasing, the solubility of chitosan decreases, while the emulsion remains stable because of oil phase droplets adsorbed on chitosan particles. If acidity increases back chitosan passes into a soluble form desorbing a dispersed oil droplets and the emulsion is reversibly separating [77]. The emulsifying ability of chitosan largely depends on the DD and MW and increased for low MW chitosan with DD less than 60% or more than 86%, whereas when the values of DD from 65% to 77%, that properties are significantly dependent on concentration [78]. Most of the practical studies on chitosan applications as an emulsifier consider it as part of complex compositions containing other surfactants. Presence both chitosan and protein emulsifiers makes a heterogeneous system more stable. Incorporation of chitosan with soybean isolate protein can increase the digestibility and stability of emulsified carotenoids [79]. The complex containing chitosan with modified  $\beta$ -lactoglobulin fibers stabilizes the fish oil emulsion in water [80]. Also, stable Pickering emulsions with corn oil are formed as a result of electrostatic interaction of chitosan with gelatin [81]. The ability of chitosan to form micelles in aqueous solutions may be used for unstable compounds as carotenoids [82] and anthocyanins [83] sustainability increasing.

*Applications of chitosan as a flocculant*

Polycationite chitosan properties can be used for drinks clarification from solid suspended particles comprising polyphenols, proteins, polysaccharides and mineral compounds [5]. Chitosan can be effectively used to clarify fruit juices [84–87], fruit [88] and grape [89] wines, beer [90] and also at the preliminary drinking water purification since this increases suspended particles sedimentation rate and increases the degree of separation of bacteria and viruses [91].

*Chitosan application in the functional foods production*

Chitosan may be used as a carrier for encapsulating and controlled delivery of probiotic ingredients in functional food products because of its high biocompatibility, emulsifying ability, polycationicity and hypoallergenic properties [92]. Components encapsulation used to prevent chemical changes during the preparation process or stomach acidic media, for lipophilic components as vitamins or flavonoids with polymer matrix delivery, to control nutraceuticals release and to mask smell and taste of some substances. The most common method of nutraceuticals encapsulation is ion gelation [6] using polyanionites – tripolyphosphate or proteins as gelling agents. Particle size depends on chitosan properties and increases with a low DD and MW associated with a decrease in the zeta potential, determined by unsaturated amino groups of the macromolecule interacting with polyanionite [93]. Can be obtained nanoparticles containing vitamins C [94,95] and B2 [96], dyes as curcumin [97] and lutein [98], flavonoids, including catechins [99,100,101] and quercetin [102], bioactive minerals selenium [103,104] and zinc [105].

A promising direction of nanoencapsulation of biologically active substances is chitosan-based multilayer composite coatings development. In this way is possible to obtain orally digestible insulin [106], and keep stable vitamin D3 in food products for up to 60 days [107].

Well known that chitosan itself exhibits biological activity. It binds fats, cholesterol and bile salt due to both hydrophobic interactions and the formation of hydrogen bonds, and as a result of electrostatic attraction of positively charged amino groups and negatively charged carboxyl groups of fatty acids.

The binding of cholesterol to chitosan was confirmed by a number of clinical studies, so chitosan may be used in the hypercholesterolemia treatment [108, 109] and included in the formulations of special foods with an anticholesterol effect. The fat binding ability of chitosan can be used in the production of dietary foods reducing the rate of lipids absorption and digestion [110]. Moreover, a composite film coating containing chitosan, gelatin and gallic acid can be used to simulate fat in food products, acting as a fat substitute by imparting organoleptic surface properties [111].

Chitoooligosaccharides have antitumor activity. Despite the large number of clinical studies, antitumor mechanism still not well understood, however, the most important factors in the action of chitosan are cancer cells viability reducing associated with the electrostatic interaction between chitosan and cell wall glycoproteins, inhibition of angiogenesis, which is the main reason for the rapid growth of cancer tumors, also a complex immunostimulating effect is possible [112].

**3. Conclusion**

A global trends analysis in the research of chitosan in food technology applications shows that the most attention to this problem is paid in Asian countries that have an abundant raw material base for its production. A large number of developments are currently proceeded in China, unlike Japan which lost its past leadership in this direction. Widespread research is also carries out in India, Spain and Iran. It should be noted that in Russia, unlike the 90s, the amount of studies on the issue has significantly decreased.

In recent years most intensive researches focused on the use of chitosan applications to extend the shelf life of foods, and also as an emulsifier, flocculant and dietary supplement. Today the explosively developing area of chitosan applying is the edible films producing. Edible coatings perform not only the function of food primary packaging, but also prevent moisture loss and oxygen access, have antimicrobial and antioxidant activity due to the unique properties of chitosan. At the same time is possible to enrich the products with additional functional ingredients as natural plant extracts, essential oils, vitamins and antimicrobial components. Thoset components enhancing the properties of the films obtained up to reach a synergistic effect over prolonging the foods shelf life. The latest trends of chitosan application are associated with freshness indicating coatings development, with vitamins, nutrients and probiotics encapsulation and delivery, and also with the biodegradable packaging engineering.

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**REFERENCES**

1. Philibert, T., Lee, B. H., Fabien, N. (2017). Current Status and New Perspectives on Chitin and Chitosan as Functional Biopolymers. *Applied Biochemistry and Biotechnology*, 181(4), 1314–1337. <https://doi.org/10.1007/s12010-016-2286-2>
2. Novinyuk, L.V., Kulyov, D.K., Negrutza, I.V., Velinzon, P.Z. (2018) Chitin- and Chitosan Biosorbents from Citric Acid Mycelial Waste. *Food systems*, 1(2), 55–62. <https://doi.org/10.21323/2618-9771-2018-1-2-55-62> (in Russian)
3. Tylliszczak, B., Drabczyk, A., Kudłacik-Kramarczyk, S., Sobczak-Kupiec, A. (2019). Sustainable Production of Chitosan. Chapter in book: Sustainable Production: Novel Trends in Energy, Environment and Material Systems, 45–60. [https://doi.org/10.1007/978-3-030-11274-5\\_4](https://doi.org/10.1007/978-3-030-11274-5_4)
4. Wang, J., Chen, C. (2014). Chitosan-based biosorbents: Modification and application for biosorption of heavy metals and radionuclides. *Bioresource Technology*, 160, 129–141. <https://doi.org/10.1016/j.biortech.2013.12.110>
5. Rinaudo, M. (2006). Chitin and chitosan: Properties and applications. *Progress in Polymer Science*, 31(7), 603–632. <https://doi.org/10.1016/j.progpolymsci.2006.06.001>

6. Gutiérrez, T. J. (2017). Chitosan Applications for the Food Industry. Chapter 8 in book: Chitosan: Derivatives, Composites and Applications, 183–232. <https://doi.org/10.1002/9781119364849.ch8>
7. Rocha, M. A. M., Coimbra, M. A., Nunes, C. (2017). Applications of chitosan and their derivatives in beverages: a critical review. *Current Opinion in Food Science*, 15, 61–69. <https://doi.org/10.1016/j.cofs.2017.06.008>
8. Ngo, D.-H., Vo, T.-S., Ngo, D.-N., Kang, K.-H., Je, J.-Y., Pham, H. N.-D., Buyn, H.-G., Kim, S.-K. (2015). Biological effects of chitosan and its derivatives. *Food Hydrocolloids*, 51, 200–216. <https://doi.org/10.1016/j.foodhyd.2015.05.023>
9. Friedman, M., Juneja, V. K. (2010). Review of Antimicrobial and Antioxidative Activities of Chitosans in Food. *Journal of Food Protection*, 73(9), 1737–1761. <https://doi.org/10.4315/0362-028x-73.9.1737>
10. Verlee, A., Mincke, S., Stevens, C. V. (2017). Recent developments in antibacterial and antifungal chitosan and its derivatives. *Carbohydrate Polymers*, 164, 268–283. <https://doi.org/10.1016/j.carbpol.2017.02.001>
11. Palma-Guerrero, J., Lopez-Jimenez, J. A., Pérez-Berná, A. J., Huang, I.-C., Jansson, H.-B., Salinas, J., Villalán, J., Read, N. D., Lopez-Llorca, L. V. (2010). Membrane fluidity determines sensitivity of filamentous fungi to chitosan. *Molecular Microbiology*, 75(4), 1021–1032. <https://doi.org/10.1111/j.1365-2958.2009.07039.x>
12. Romanazzi, G., Feliziani, E., Baños, S. B., Sivakumar, D. (2015). Shelf life extension of fresh fruit and vegetables by chitosan treatment. *Critical Reviews in Food Science and Nutrition*, 57(3), 579–601. <https://doi.org/10.1080/10408398.2014.900474>
13. Hassan, B., Chatha, S. A. S., Hussain, A. I., Zia, K. M., Akhtar, N. (2018). Recent advances on polysaccharides, lipids and protein based edible films and coatings: A review. *International Journal of Biological Macromolecules*, 109, 1095–1107. <https://doi.org/10.1016/j.ijbiomac.2017.11.097>
14. Mujtaba, M., Morsi, R. E., Kerch, G., Elsabee, M. Z., Kaya, M., Labidi, J., Khawar, K. M. (2019). Current advancements in chitosan-based film production for food technology; A review. *International Journal of Biological Macromolecules*, 121, 889–904. <https://doi.org/10.1016/j.ijbiomac.2018.10.109>
15. Kerch, G., Korkhov, V. (2010). Effect of storage time and temperature on structure, mechanical and barrier properties of chitosan-based films. *European Food Research and Technology*, 232(1), 17–22. <https://doi.org/10.1007/s00217-010-1356-x>
16. Leceta, I., Molinaro, S., Guerrero, P., Kerry, J. P., de la Caba, K. (2015). Quality attributes of map packaged ready-to-eat baby carrots by using chitosan-based coatings. *Postharvest Biology and Technology*, 100, 142–150. <https://doi.org/10.1016/j.postharvbio.2014.09.022>
17. Villafañe, F. (2016). Edible coatings for carrots. *Food Reviews International*, 33(1), 84–103. <https://doi.org/10.1080/87559129.2016.1150291>
18. Leandro, D. S. P., Bitencourt, T. A., Saltoratto, A. L., Selegim, M. H., Assis, O. B. (2018). Antifungal activity of chitosan and its quaternized derivative in gel form and as an edible coating on cut cherry tomatoes. *Journal of Agricultural Sciences*, 63(3), 271–285. <https://doi.org/10.2298/jas1803271s>
19. Olawuyi, I. F., Lee, W. (2019). Influence of chitosan coating and packaging materials on the quality characteristics of fresh-cut cucumber. *Korean Journal of Food Preservation*, 26(4), 371–380. <https://doi.org/10.11002/kjfp.2019.26.4.371>
20. Ali, A., Noh, N. M., Mustafa, M. A. (2015). Antimicrobial activity of chitosan enriched with lemongrass oil against anthracnose of bell pepper. *Food Packaging and Shelf Life*, 3, 56–61. <https://doi.org/10.1016/j.fpsl.2014.10.005>
21. Zahoorullah, S. M., Dakshayani, L., Rani, A. S., Venkateswerlu, G. (2017). Effect of Chitosan Coating on the Post Harvest Quality of Banana during Storage. *Asian Journal of Biotechnology and Bioresource Technology*, 1(1), 1–10. <https://doi.org/10.9734/ajbt/2017/34732>
22. Li, H., Wang, Y., Liu, F., Yang, Y., Wu, Z., Cai, H., Zhang, Q., Wang, Y., Li, P. (2015). Effects of chitosan on control of postharvest blue mold decay of apple fruit and the possible mechanisms involved. *Scientia Horticulturae*, 186, 77–83. <https://doi.org/10.1016/j.scienta.2015.02.014>
23. Varasteh, F., Arzani, K., Barzegar, M., Zamani, Z. (2017). Pomegranate (Punica granatum L.) Fruit Storability Improvement Using Pre-storage Chitosan Coating Technique. *Journal of Agricultural Science Technology*, 19(2), 389–400.
24. Wang, J., Yang, B., Zhang, S., Cao, J., Jiang, W. (2016). Effect of thymol on antifungal ability of chitosan coating against Penicillium expansum in Yali pear. *Emirates Journal of Food and Agriculture*, 28(10), 725–731. <https://doi.org/10.9755/ejfa.2015-09-788>
25. Kumar, P., Sethi, S., Sharma, R. R., Srivastav, M., Varghese, E. (2017). Effect of chitosan coating on postharvest life and quality of plum during storage at low temperature. *Scientia Horticulturae*, 226, 104–109. <https://doi.org/10.1016/j.scienta.2017.08.037>
26. Badawy, M. E. I., Rabea, E. I., El-Nouby, M. A. M., Ismail, R. I. A., Taktak, N. E. M., (2016). Strawberry Shelf Life, Composition, and Enzymes Activity in Response to Edible Chitosan Coatings. *International Journal of Fruit Science*, 17(2), 117–136. <https://doi.org/10.1080/15538362.2016.1219290>
27. Bhanushree, L. S., Vasudeva, K. R., Suresha, G. J., Sadananda, G. K., Mohamad Tayeebulla, H., Halesh, G. K. (2018). Influence of chitosan on postharvest behavior of papaya (Carica papaya L.) Fruits under different storage conditions. *Journal of Pharmacognosy and Phytochemistry*, 7(2), 2010–2014.
28. Jongsri, P., Wangsomboondee, T., Rojsitthisak, P., Seraypheap, K. (2016). Effect of molecular weights of chitosan coating on postharvest quality and physicochemical characteristics of mango fruit. *LWT*, 73, 28–36. <https://doi.org/10.1016/j.lwt.2016.05.038>
29. Remya, S., Mohan, C. O., Bindu, J., Sivaraman, G. K., Venkateshwarlu, G., Ravishankar, C. N. (2015). Effect of chitosan based active packaging film on the keeping quality of chilled stored barracuda fish. *Journal of Food Science and Technology*, 53(1), 685–693. <https://doi.org/10.1007/s13197-015-2018-6>
30. Chang, W., Liu, F., Sharif, H. R., Huang, Z., Goff, H. D., Zhong, F. (2018). Preparation of chitosan films by neutralization for improving their preservation effects on chilled meat. *Food Hydrocolloids*, 90, 50–61. <https://doi.org/10.1016/j.foodhyd.2018.09.026>
31. Chantarasataporn, P., Tepkasikul, P., Kingcha, Y., Yoksan, R., Pichyangkura, R., Visessanguan, W., Chirachanchai, S. (2014). Water-based oligochitosan and nanowhisker chitosan as potential food preservatives for shelf-life extension of minced pork. *Food Chemistry*, 159, 463–470. <https://doi.org/10.1016/j.foodchem.2014.03.019>
32. Tayel, A. A., Ibrahim, S. I. A., Al-Saman, M. A., Moussa, S. H. (2014). Production of fungal chitosan from date wastes and its application as a biopreservative for minced meat. *International Journal of Biological Macromolecules*, 69, 471–475. <https://doi.org/10.1016/j.ijbiomac.2014.05.072>
33. Lozano-Navarro, J., Díaz-Zavala, N., Velasco-Santos, C., Melo-Banda, J., Páramo-García, U., Paraguay-Delgado, F., Martínez-Hernández, A. L., Zapién-Castillo, S. (2018). Chitosan-Starch Films with Natural Extracts: Physical, Chemical, Morphological and Thermal Properties. *Materials*, 11(1), 120. <https://doi.org/10.3390/ma11010120>
34. Shariatinia, Z., Fazli, M. (2015). Mechanical properties and antibacterial activities of novel nanobiocomposite films of chitosan and starch. *Food Hydrocolloids*, 46, 112–124. <https://doi.org/10.1016/j.foodhyd.2014.12.026>
35. Sundaram, J., Pant, J., Goudie, M. J., Mani, S., Handa, H. (2016). Antimicrobial and Physicochemical Characterization of Biodegradable, Nitric Oxide-Releasing Nanocellulose–Chitosan Packaging Membranes. *Journal of Agricultural and Food Chemistry*, 64(25), 5260–5266. <https://doi.org/10.1021/acs.jafc.6b01936>
36. Bansal, M., Chauhan, G. S., Kaushik, A., Sharma, A. (2016). Extraction and functionalization of bagasse cellulose nanofibres to Schiff-base based antimicrobial membranes. *International Journal of Biological Macromolecules*, 91, 887–894. <https://doi.org/10.1016/j.ijbiomac.2016.06.045>
37. Poverenov, E., Danino, S., Horev, B., Granit, R., Vinokur, Y., Rodov, V. (2013). Layer-by-Layer Electrostatic Deposition of Edible Coating on Fresh Cut Melon Model: Anticipated and Unexpected Effects of Alginate–Chitosan Combination. *Food and Bioprocess Technology*, 7(5), 1424–1432. <https://doi.org/10.1007/s11947-013-1134-4>
38. Souza, M. P.; Vaz, A. F. M.; Cerqueira, M. A.; Texeira, J. A.; Vicente, A. A.; Carneiro-da-Cunha, M. G. (2015). Effect of an ediblenanomultilayer coating by electrostatic self-assembly on the shelf life of fresh-cut mangoes. *Food Bioprocess Technology*, 8(3), 647–654. <https://doi.org/10.1007/s11947-014-1436-1>
39. Younis, H. G. R., Zhao, G. (2019). Physicochemical properties of the edible films from the blends of high methoxyl apple pectin and chitosan. *International Journal of Biological Macromolecules*, 131, 1057–1066. <https://doi.org/10.1016/j.ijbiomac.2019.03.096>
40. Volpe, S., Torriero, E., Cavella, S. (2017). Use of Chitosan and Chitosan-Caseinate Coating to Prolong Shelf Life of Minimally Processed Apples. *SLIM 2017 – Shelf-life International Meeting – Special issue of Italian Journal of Food Science*, 29(5), 30–35.
41. Pérez Córdoba, L. J., Sobral, P. J. A. (2017). Physical and antioxidant properties of films based on gelatin, gelatin-chitosan or gelatin-sodium caseinate blends loaded with nanoemulsified active compounds. *Journal of Food Engineering*, 213, 47–53. <https://doi.org/10.1016/j.jfoodeng.2017.05.023>
42. Qiao, C., Ma, X., Zhang, J., Yao, J. (2017). Molecular interactions in gelatin/chitosan composite films. *Food Chemistry*, 235, 45–50. <https://doi.org/10.1016/j.foodchem.2017.05.045>
43. Ahmed, S., Ikram, S. (2016). Chitosan and gelatin based biodegradable packaging films with UV-light protection. *Journal of Photochemistry and Photobiology B: Biology*, 163, 115–124. <https://doi.org/10.1016/j.jphoto-biol.2016.08.023>
44. Ahmad, M., Nirmal, N. P., Danish, M., Chuprom, J., Jafarzedeh, S. (2016). Characterisation of composite films fabricated from collagen/chitosan and collagen/soy protein isolate for food packaging applications. *RSC Advances*, 6(85), 82191–82204. <https://doi.org/10.1039/c6ra13043g>
45. Velickova, E., Winkelhausen, E., Kuzmanova, S., Moldão-Martins, M., Alves, V. D. (2013). Characterization of multilayered and composite edible films from chitosan and beeswax. *Food Science and Technology International*, 21(2), 83–93. <https://doi.org/10.1177/1082013213511807>
46. Wang, H., Qian, J., Ding, F. (2018). Emerging Chitosan-Based Films for Food Packaging Applications. *Journal of Agricultural and Food Chemistry*, 66(2), 395–413. <https://doi.org/10.1021/acs.jafc.7b04528>
47. Dominguez-Martinez, B. M., Martínez-Flores, H. E., Berrios, J. D. J., Otoni, C. G., Wood, D. F., Velazquez, G. (2016). Physical Characterization of Biodegradable Films Based on Chitosan, Polyvinyl Alcohol and Opuntia Mucilage. *Journal of Polymers and the Environment*, 25(3), 683–691. <https://doi.org/10.1007/s10924-016-0851-y>
48. Yang, W., Gmczarek, J. S., Fortunati, E., Kozanecki, M., Mazzaglia, A., Balestra, G. M., Kenny, J. M., Torre, L., Puglia, D. (2016). Antioxidant and antibacterial lignin nanoparticles in polyvinyl alcohol/chitosan films for

active packaging. *Industrial Crops and Products*, 94, 800–811. <https://doi.org/10.1016/j.indcrop.2016.09.061>

49. Talón, E., Trifkovic, K. T., Nedovic, V. A., Bugarski, B. M., Vargas, M., Chiralt, A., González-Martínez, C. (2017). Antioxidant edible films based on chitosan and starch containing polyphenols from thyme extracts. *Carbohydrate Polymers*, 157, 1153–1161. <https://doi.org/10.1016/j.carbpol.2016.10.080>
50. Kalaycıoğlu, Z., Torlak, E., Akin-Evingür, G., Özen, İ., Erim, F. B. (2017). Antimicrobial and physical properties of chitosan films incorporated with turmeric extract. *International Journal of Biological Macromolecules*, 101, 882–888. <https://doi.org/10.1016/j.ijbiomac.2017.03.174>
51. Beigzadeh Ghelejlou, S., Esmailli, M., Almasi, H. (2016). Characterization of chitosan–nanoclay bionanocomposite active films containing milk thistle extract. *International Journal of Biological Macromolecules*, 86, 613–621. <https://doi.org/10.1016/j.ijbiomac.2016.02.012>
52. Kaya, M., Ravikumar, P., Ilk, S., Mujtaba, M., Akyuz, L., Labidi, J., Salaberria, A. M., Cakmak, Y. S., Erkul, S. K. (2018). Production and characterization of chitosan based edible films from *Berberis crataegina*'s fruit extract and seed oil. *Innovative Food Science and Emerging Technologies*, 45, 287–297. <https://doi.org/10.1016/j.ifset.2017.11.013>
53. Bonilla, J., Sobral, P. J. A. (2016). Investigation of the physicochemical, antimicrobial and antioxidant properties of gelatin–chitosan edible film mixed with plant ethanolic extracts. *Food Bioscience*, 16, 17–25. <https://doi.org/10.1016/j.fbio.2016.07.003>
54. Tan, Y. M., Lim, S. H., Tay, B. Y., Lee, M. W., Thian, E. S. (2015). Functional chitosan–based grapefruit seed extract composite films for applications in food packaging technology. *Materials Research Bulletin*, 69, 142–146. <https://doi.org/10.1016/j.materresbull.2014.11.041>
55. Kumar-Krishnan, S., Prokhorov, E., Hernández-Iturriaga, M., Mota-Morales, J. D., Vázquez-Lepe, M., Kovalenko, Yu., Sanchez, I.C., Luna-Bárceñas, G. (2015). Chitosan/silver nanocomposites: Synergistic antibacterial action of silver nanoparticles and silver ions. *European Polymer Journal*, 67, 242–251. <https://doi.org/10.1016/j.eurpolymj.2015.03.066>
56. Raghavendra, G. M., Jung, J., Kim, D., Seo, J. (2016). Microwave assisted antibacterial chitosan–silver nanocomposite films. *International Journal of Biological Macromolecules*, 84, 281–288. <https://doi.org/10.1016/j.ijbiomac.2015.12.026>
57. Yu, W.-Z., Zhang, Y., Liu, X., Xiang, Y., Li, Z., Wu, S. (2018). Synergistic antibacterial activity of multi components in lysozyme/chitosan/silver/hydroxyapatite hybrid coating. *Materials Design*, 139, 351–362. <https://doi.org/10.1016/j.matdes.2017.11.018>
58. Al-Naamani, L., Dobretsov, S., Dutta, J. (2016). Chitosan–zinc oxide nanoparticle composite coating for active food packaging applications. *Innovative Food Science and Emerging Technologies*, 38, 231–237. <https://doi.org/10.1016/j.ifset.2016.10.010>
59. Al-Naamani, L., Dobretsov, S., Dutta, J., Burgess, J. G. (2017). Chitosan–zinc oxide nanocomposite coatings for the prevention of marine biofouling. *Chemosphere*, 168, 408–417. <https://doi.org/10.1016/j.chemosphere.2016.10.033>
60. Rahman, P. M., Mujeeb, V. M. A., Muraleedharan, K. (2017). Flexible chitosan–nano ZnO antimicrobial pouches as a new material for extending the shelf life of raw meat. *International Journal of Biological Macromolecules*, 97, 382–391. <https://doi.org/10.1016/j.ijbiomac.2017.01.052>
61. Can, Ö. P., Yalcin, H., Arslan, A. (2018). Effects of chitosan coating and rosemary oil on rainbow trout (*Oncorhynchus mykiss*, W. 1792) filets. *Indian Journal of Animal Research*, 52(1), 160–166. <https://doi.org/10.18805/ijar.v0i0f.6820>
62. Valipour Kootenaie, F., Ariaii, P., Khademi Shurmasti, D., Nemati, M. (2016). Effect of Chitosan Edible Coating Enriched with Eucalyptus Essential Oil and  $\alpha$ -Tocopherol on Silver Carp Fillets Quality During Refrigerated Storage. *Journal of Food Safety*, 37(1), e12295. <https://doi.org/10.1111/jfs.12295>
63. Xu, T., Gao, C., Feng, X., Huang, M., Yang, Y., Shen, X., Tang, X. (2019). Cinnamon and clove essential oils to improve physical, thermal and antimicrobial properties of chitosan–gum arabic polyelectrolyte complexed films. *Carbohydrate Polymers*, 217, 116–125. <https://doi.org/10.1016/j.carbpol.2019.03.084>
64. Noshirvani, N., Ghanbarzadeh, B., Gardrat, C., Rezaei, M. R., Hashemi, M., Le Coz, C., Coma, V. (2017). Cinnamon and ginger essential oils to improve antifungal, physical and mechanical properties of chitosan–carboxymethyl cellulose films. *Food Hydrocolloids*, 70, 36–45. <https://doi.org/10.1016/j.foodhyd.2017.03.015>
65. Wang, Y., Xia, Y., Zhang, P., Ye, L., Wu, L., He, S. (2016). Physical Characterization and Pork Packaging Application of Chitosan Films Incorporated with Combined Essential Oils of Cinnamon and Ginger. *Food and Bioprocess Technology*, 10(5), 503–511. <https://doi.org/10.1007/s11947-016-1833-8>
66. Shao, X., Cao, B., Xu, F., Xie, S., Yu, D., Wang, H. (2015). Effect of post-harvest application of chitosan combined with clove oil against citrus green mold. *Postharvest Biology and Technology*, 99, 37–45. <https://doi.org/10.1016/j.postharvbio.2014.07.014>
67. Ospina, J. D., Grande, C. D., Monsalve, L. V., Advíncula, R. C., Mina, J. H., Valencia, M. E., Fan, J., Rodrigues, D. (2019). Evaluation of the chitosan films of essential oils from *Origanum vulgare* L. (oregano) and *Rosmarinus officinalis* L. (rosemary). *Revista Cubana de Plantas Medicinales* [Online], 24(1). [Electronic resource: <http://revplantasmedicinales.sld.cu/index.php/pla/article/view/655/355> / Access date 18.02.2020 r.]
68. Quesada, J., Sendra, E., Navarro, C., Sayas-Barberá, E. (2016). Antimicrobial Active Packaging including Chitosan Films with Thymus vulgaris L. Essential Oil for Ready-to-Eat Meat. *Foods*, 5(4), 57. <https://doi.org/10.3390/foods5030057>
69. Sharafati Chaleshtori, F., Taghizadeh, M., Rafeian-kopaei, M., Sharafati-chaleshtori, R. (2015). Effect of Chitosan Incorporated with Cumin and Eucalyptus Essential Oils As Antimicrobial Agents on Fresh Chicken Meat. *Journal of Food Processing and Preservation*, 40(3), 396–404. <https://doi.org/10.1111/jfpp.12616>
70. Sharafati Chaleshtori, F., Sharafati Chaleshtori, R. (2017). Antimicrobial activity of chitosan incorporated with lemon and oregano essential oils on broiler breast meat during refrigerated storage. *Nutrition and Food Science*, 47(5), 306–317. <https://doi.org/10.1108/nfs-08-2016-0123>
71. Perdones, Á., Escriche, I., Chiralt, A., Vargas, M. (2016). Effect of chitosan–lemon essential oil coatings on volatile profile of strawberries during storage. *Food Chemistry*, 197, 979–986. <https://doi.org/10.1016/j.foodchem.2015.11.054>
72. Silva-Pereira, M. C., Teixeira, J. A., Pereira-Júnior, V. A., Stefani, R. (2015). Chitosan/corn starch blend films with extract from *Brassica oleracea* (red cabbage) as a visual indicator of fish deterioration. *LWT – Food Science and Technology*, 61(1), 258–262. <https://doi.org/10.1016/j.lwt.2014.11.041>
73. Ma, Q., Liang, T., Cao, L., Wang, L. (2018). Intelligent poly (vinyl alcohol)-chitosan nanoparticles–mulberry extracts films capable of monitoring pH variations. *International Journal of Biological Macromolecules*, 108, 576–584. <https://doi.org/10.1016/j.ijbiomac.2017.12.049>
74. Cazón, P., Vázquez, M., Velázquez, G. (2018). Novel composite films based on cellulose reinforced with chitosan and polyvinyl alcohol: Effect on mechanical properties and water vapour permeability. *Polymer Testing*, 69, 536–544. <https://doi.org/10.1016/j.polymertesting.2018.06.016>
75. Fathima, P. E., Panda, S. K., Ashraf, P. M., Varghese, T. O., Bindu, J. (2018). Polylactic acid/chitosan films for packaging of Indian white prawn (*Penaeus indicus*). *International Journal of Biological Macromolecules*, 117, 1002–1010. <https://doi.org/10.1016/j.ijbiomac.2018.05.214>
76. Pal, A. K., Katiyar, V. (2016). Nanoamphiphilic Chitosan Dispersed Poly(lactic acid) Bionanocomposite Films with Improved Thermal, Mechanical, and Gas Barrier Properties. *Biomacromolecules*, 17(8), 2603–2618. <https://doi.org/10.1021/acs.biomac.6b00619>
77. Liu, H., Wang, C., Zou, S., Wei, Z., Tong, Z. (2012). Simple, Reversible Emulsion System Switched by pH on the Basis of Chitosan without Any Hydrophobic Modification. *Langmuir*, 28(30), 11017–11024. <https://doi.org/10.1021/la3021113>
78. Rodríguez, M.S., Albertengo, L.A., Agulló, E. (2002). Emulsification capacity of chitosan. *Carbohydrate Polymers*, 48(3), 271–276. [https://doi.org/10.1016/s0144-8617\(01\)00258-2](https://doi.org/10.1016/s0144-8617(01)00258-2)
79. Zhang, C., Xu, W., Jin, W., Shah, B. R., Li, Y., Li, B. (2015). Influence of anionic alginate and cationic chitosan on physicochemical stability and carotenoids bioaccessibility of soy protein isolate-stabilized emulsions. *Food Research International*, 77, 419–425. <https://doi.org/10.1016/j.foodres.2015.09.020>
80. Chang, H. W., Tan, T. B., Tan, P. Y., Nehdi, I. A., Sbihi, H. M., Tan, C. P. (2019). Microencapsulation of Fish Oil-In-Water Emulsion Using Thiol-Modified  $\beta$ -Lactoglobulin Fibrils-Chitosan Complex. *Journal of Food Engineering*. <https://doi.org/10.1016/j.jfoodeng.2019.07.027>
81. Wang, X.-Y., Heuzey, M.-C. (2016). Pickering emulsion gels based on insoluble chitosan/gelatin electrostatic complexes. *RSC Advances*, 6(92), 89776–89784. <https://doi.org/10.1039/c6ra10378b>
82. Zhang, C., Xu, W., Jin, W., Shah, B. R., Li, Y., Li, B. (2015). Influence of anionic alginate and cationic chitosan on physicochemical stability and carotenoids bioaccessibility of soy protein isolate-stabilized emulsions. *Food Research International*, 77, 419–425. <https://doi.org/10.1016/j.foodres.2015.09.020>
83. He, B., Ge, J., Yue, P., Yue, X., Fu, R., Liang, J., Gao, X. (2017). Loading of anthocyanins on chitosan nanoparticles influences anthocyanin degradation in gastrointestinal fluids and stability in a beverage. *Food Chemistry*, 221, 1671–1677. <https://doi.org/10.1016/j.foodchem.2016.10.120>
84. Abdelmalek, B. E., Sila, A., Haddar, A., Bougataf, A., Ayadi, M. A. (2017).  $\beta$ -Chitin and chitosan from squid gladius: Biological activities of chitosan and its application as clarifying agent for apple juice. *International Journal of Biological Macromolecules*, 104, 953–962. <https://doi.org/10.1016/j.ijbiomac.2017.06.107>
85. Taştan, Ö., Baysal, T. (2017). Chitosan as a novel clarifying agent on clear apple juice production: Optimization of process conditions and changes on quality characteristics. *Food Chemistry*, 237, 818–824. <https://doi.org/10.1016/j.foodchem.2017.06.025>
86. Lei, W., Li, C., Wang, N., Ji, T., Fu, X. (2016). Study on Clarification Effect of Chitosan on Cantaloupe Juice. *The Food Industry*, 8, 11–13. (in Chinese)
87. Taştan, Ö., Baysal, T. (2015). Clarification of pomegranate juice with chitosan: Changes on quality characteristics during storage. *Food Chemistry*, 180, 211–218. <https://doi.org/10.1016/j.foodchem.2015.02.053>
88. Jiang, Y., You, T., Liu, J., Bao, H. (2015). Application of Chitosan Flocculation on Clarifying of Traditional Chinese Medicine and Fruit Wine. *The Food Industry*, 2, 228–231. (in Chinese)
89. Qi, G., Gao, D. (2016). Clarification of Dry Red Wine by Chitosan. *Liquor-Making Science & Technology*, 10, 27–29. (in Chinese)

90. Gassara, F., Antzak, C., Ajila, C. M., Sarma, S. J., Brar, S. K., Verma, M. (2015). Chitin and chitosan as natural flocculants for beer clarification. *Journal of Food Engineering*, 166, 80–85. <https://doi.org/10.1016/j.jfoodeng.2015.05.028>
91. Abebe, L., Chen, X., Sobsey, M. (2016). Chitosan Coagulation to Improve Microbial and Turbidity Removal by Ceramic Water Filtration for Household Drinking Water Treatment. *International Journal of Environmental Research and Public Health*, 13(5), 269. <https://doi.org/10.3390/ijerph13030269>
92. Akbari-Alavijeh, S., Shaddel, R., Jafari, S. M. (2019). Nanostructures of chitosan for encapsulation of food ingredients. Chapter in Book: *Bio-polymer Nanostructures for Food Encapsulation Purposes*, 381–418. Academic Press <https://doi.org/10.1016/b978-0-12-815663-6.00014-8>
93. Al-Nemrawi, N. K., Alsharif, S. S. M., Dave, R. H. (2018). Preparation of chitosan-TPP nanoparticles: the influence of chitosan polymeric properties and formulation variables. *International Journal of Applied Pharmaceutics*, 10(5), 60–65. <https://doi.org/10.22159/ijap.2018v10i5.26375>
94. Jiménez-Fernández, E., Ruyra, A., Roher, N., Zuasti, E., Infante, C., Fernández-Díaz, C. (2014). Nanoparticles as a novel delivery system for vitamin C administration in aquaculture. *Aquaculture*, 432, 426–433. <https://doi.org/10.1016/j.aquaculture.2014.03.006>
95. Alishahi, A., Mirvaghefi, A., Tehrani, M. R., Farahmand, H., Shojaosadati, S. A., Dorkoosh, F. A., Elsabee, M. Z. (2011). Shelf life and delivery enhancement of vitamin C using chitosan nanoparticles. *Food Chemistry*, 126(3), 935–940. <https://doi.org/10.1016/j.foodchem.2010.11.086>
96. Azevedo, M. A., Bourbon, A. I., Vicente, A. A., Cerqueira, M. A. (2014). Alginate/chitosan nanoparticles for encapsulation and controlled release of vitamin B2. *International Journal of Biological Macromolecules*, 71, 141–146. <https://doi.org/10.1016/j.ijbiomac.2014.05.036>
97. Vishwakarma, A., Sriram, P., Preetha, S. P., Tirumurugaan, K. G., Nagarajan, K., Pandian, K. (2019). Synthesis and characterization of Chitosan/TPP encapsulated curcumin nanoparticles and its antibacterial efficacy against colon bacteria. *International Journal of Chemical Studies*, 7(3), 602–606.
98. Arunkumar, R., Harish Prashanth, K. V., Baskaran, V. (2013). Promising interaction between nanoencapsulated lutein with low molecular weight chitosan: Characterization and bioavailability of lutein in vitro and in vivo. *Food Chemistry*, 141(1), 327–337. <https://doi.org/10.1016/j.foodchem.2013.02.108>
99. Dube, A., Nicolazzo, J. A., Larson, I. (2011). Chitosan nanoparticles enhance the plasma exposure of (–)-epigallocatechin gallate in mice through an enhancement in intestinal stability. *European Journal of Pharmaceutical Sciences*, 44(3), 422–426. <https://doi.org/10.1016/j.ejps.2011.09.004>
100. Hu, B., Ting, Y., Yang, X., Tang, W., Zeng, X., Huang, Q. (2012). Nanochemoprevention by encapsulation of (–)-epigallocatechin-3-gallate with bioactive peptides/chitosan nanoparticles for enhancement of its bioavailability. *Chemical Communications*, 48(18), 2421. <https://doi.org/10.1039/c2cc17295j>
101. Hu, B., Ting, Y., Zeng, X., Huang, Q. (2012). Cellular uptake and cytotoxicity of chitosan-caseinophosphopeptides nanocomplexes loaded with epigallocatechin gallate. *Carbohydrate Polymers*, 89(2), 362–370. <https://doi.org/10.1016/j.carbpol.2012.03.015>
102. Zhang, Y., Yang, Y., Tang, K., Hu, X., Zou, G. (2008). Physicochemical characterization and antioxidant activity of quercetin-loaded chitosan nanoparticles. *Journal of Applied Polymer Science*, 107(2), 891–897. <https://doi.org/10.1002/app.26402>
103. Zhang, S., Luo, Y., Zeng, H., Wang, Q., Tian, F., Song, J., Cheng, W.-H. (2011). Encapsulation of selenium in chitosan nanoparticles improves selenium availability and protects cells from selenium-induced DNA damage response. *The Journal of Nutritional Biochemistry*, 22(12), 1137–1142. <https://doi.org/10.1016/j.jnutbio.2010.09.014>
104. Luo, Y., Zhang, B., Cheng, W.-H., Wang, Q. (2010). Preparation, characterization and evaluation of selenite-loaded chitosan/TPP nanoparticles with or without zein coating. *Carbohydrate Polymers*, 82(3), 942–951. <https://doi.org/10.1016/j.carbpol.2010.06.029>
105. Deshpande, P., Dapkekar, A., Oak, M. D., Paknikar, K. M., Rajwade, J. M. (2017). Zinc complexed chitosan/TPP nanoparticles: A promising micronutrient nanocarrier suited for foliar application. *Carbohydrate Polymers*, 165, 394–401. <https://doi.org/10.1016/j.carbpol.2017.02.061>
106. Lopes, M., Shrestha, N., Correia, A., Shahbazi, M.-A., Sarmiento, B., Hirvonen, J., Veiga, F., Seica, R., Ribeiro, A., Santos, H. A. (2016). Dual chitosan/albumin-coated alginate/dextran sulfate nanoparticles for enhanced oral delivery of insulin. *Journal of Controlled Release*, 232, 29–41. <https://doi.org/10.1016/j.jconrel.2016.04.012>
107. Rabelo, R. S., Oliveira, I. F., da Silva, V. M., Prata, A. S., Hubinger, M. D. (2018). Chitosan coated nanostructured lipid carriers (NLCs) for loading Vitamin D: A physical stability study. *International Journal of Biological Macromolecules*, 119, 902–912. <https://doi.org/10.1016/j.ijbiomac.2018.07.174>
108. Ylitalo, R., Lehtinen, S., Wuolijoki, E., Ylitalo, P., Lehtimäki, T. (2002). Cholesterol-lowering properties and safety of chitosan. *Arzneimittelforschung*, 52(01), 1–7. <https://doi.org/10.1055/s-0031-1299848>
109. Bokura, H., Kobayashi, S. (2003). Chitosan decreases total cholesterol in women: a randomized, double-blind, placebo-controlled trial. *European Journal of Clinical Nutrition*, 57(5), 721–725. <https://doi.org/10.1038/sj.ejcn.1601603>
110. Patti, A. M., Katsiki, N., Nikolic, D., Al-Rasadi, K., Rizzo, M. (2014). Nutraceuticals in Lipid-Lowering Treatment. *Angiology*, 66(5), 416–421. <https://doi.org/10.1177/0003319714542999>
111. Rezaee, M., Askari, G., EmamDjomeh, Z., Salami, M. (2018). Effect of organic additives on physicochemical properties and anti-oxidant release from chitosan-gelatin composite films to fatty food simulant. *International Journal of Biological Macromolecules*, 114, 844–850. <https://doi.org/10.1016/j.ijbiomac.2018.03.122>
112. Zou, P., Yang, X., Wang, J., Li, Y., Yu, H., Zhang, Y., Liu, G. (2016). Advances in characterisation and biological activities of chitosan and chitosan oligosaccharides. *Food Chemistry*, 190, 1174–1181. <https://doi.org/10.1016/j.foodchem.2015.06.076>

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